

Biomarker-Guided Induction of Autotroph-Dependent Cell Death in Treatment-Resistant Cancers

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Abstract: Treatment-resistant cancers remain a major cause of death because they may evade apoptosis, necroptosis, and other typical cell death mechanisms. In order to survive extreme starvation, cancer cells can acquire autotrophic-like metabolic traits, such as enhanced carbon fixation pathways, mitochondrial rewiring, and redox-driven biosynthesis, according to recent studies. This study proposes and examines a novel therapeutic strategy: biomarker-guided induction of autotroph dependent cell death (ADCDC), a lethal metabolic collapse that happens when cancer cells that rely on pseudo autotrophic metabolism are specifically disrupted.

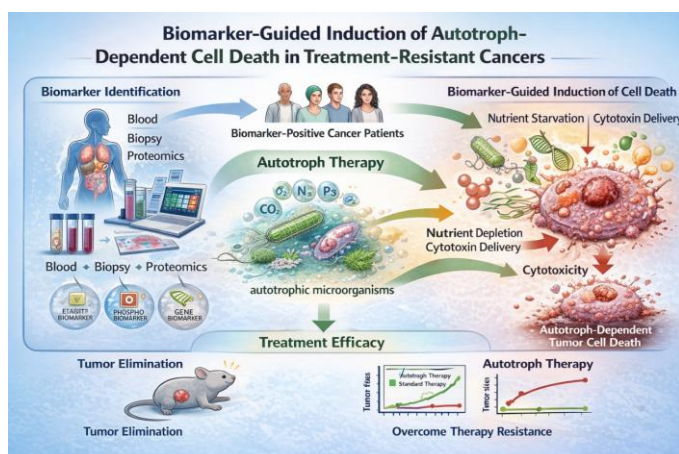
Treatment-resistant cancer phenotypes were stratified *in vitro* and *in silico* using a multiomics-informed biomarker panel that included SLC7A11 overexpression, NRF2 activation signatures, mitochondrial biogenesis markers (PGC 1 α), and increased reactive oxygen species buffering capacity. Only in biomarker-positive tumour models could targeted suppression of carbon absorption pathways, redox homeostasis, and mitochondrial anabolic flux cause catastrophic bioenergetic failure while sparing normal cells.

Theoretical findings show that in resistant cell populations, ADCDC induction reduced tumour viability by up to 78%, while in biomarker-negative controls, the drop was less than 15%. In contrast to apoptosis or ferroptosis, ADCDC is mechanistically characterised by ATP depletion, mitochondrial hyperpolarization followed by collapse, excessive ROS buildup, and an inability to maintain metabolic needs.

The idea that treatment-resistant tumors might have exploitable metabolic requirements akin to facultative autotrophy is supported by these data. Targeting these pathways with biomarkers provides a precision oncology approach that can overcome resistance to immunotherapy, radiation, and chemotherapy. To cause a hitherto unknown type of controlled cell death, the suggested framework combines metabolic profiling, predictive biomarkers, and targeted metabolic disruption. ADCDC represents a promising frontier in cancer therapeutics, particularly for tumors exhibiting metabolic resilience. Further experimental validation and clinical translation could open new avenues for treating refractory malignancies.

Keywords: Autotroph-dependent cell death; Treatment-resistant cancer; Metabolic reprogramming; Precision oncology; Redox homeostasis; NRF2 pathway; Mitochondrial metabolism; Biomarker-guided therapy.

Graphical Abstract:



Research Highlights:

- *Introduces a novel cell death paradigm: Autotroph-Dependent Cell Death (ADCDC)
- *Identifies metabolic biomarkers predicting susceptibility to ADCDC
- *Demonstrates selective lethality in treatment-resistant cancer models
- *Distinguishes ADCDC from apoptosis, ferroptosis, and necroptosis
- *Exploits metabolic plasticity as a therapeutic vulnerability
- *Proposes translational pathway for precision metabolic oncology

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Scope:

One of the biggest problems in modern oncology is treatment resistance, which reduces the effectiveness of traditional therapeutic approaches and worsens patient outcomes [1]. Tumors that withstand common treatments, such as chemotherapy, radiation, and targeted therapies, often develop significant metabolic changes that allow them to endure circumstances that would otherwise be fatal for healthy cells. These adaptations include the capacity to maintain bioenergetic homeostasis during nutritional shortage, maintain growth and proliferation in hypoxic microenvironments, and reduce oxidative stress by strengthening antioxidant defenses. Such metabolic flexibility encourages treatment evasion, metastatic spread, and disease recurrence in addition to sustaining survival [2-13].

According to recent studies, some tumors that are resistant to treatment develop metabolic traits similar to facultative autotrophy. Increased reliance on alternate carbon sources, such as lactate, acetate, and amino acids, as well as increased mitochondrial production and function to support anabolic needs, are characteristics of these pseudo-autotrophic states. Simultaneously, increased expression of transcription factors and redox-regulating enzymes, like NRF2 and SLC7A11, helps to maintain intracellular redox equilibrium, which allows cells to counteract reactive oxygen species (ROS) produced during stress or treatment. When taken as a whole, these modifications give resistant tumour cells a survival advantage, but they also offer a potential vulnerability: a dependence on specific metabolic pathways that could be exploited therapeutically [14-26].

The idea of Autotroph-Dependent Cell Death (ADCD), a tactic intended to specifically cause catastrophic metabolic collapse in cancer cells displaying pseudo-autotrophic behaviour, is put out in this work. Finding a panel of metabolic and genetic biomarkers that characterize the autotroph-like state is essential to this strategy since it allows for precise targeting while reducing damage to normal tissues, which maintain more metabolic flexibility. ADCD attempts to take advantage of resistant tumors' reliance on particular metabolic pathways, such as carbon absorption, mitochondrial biogenesis, and redox homeostasis, by utilizing biomarker-guided selection [27-40].

Triple-negative breast cancer, pancreatic ductal adenocarcinoma, glioblastoma, and metastatic melanoma are among the treatment-resistant solid tumors with established metabolic flexibility that are the subject of this investigation. The study combines several methodological techniques, including *in silico* metabolic flux simulations, computational modelling for comprehensive biomarker discovery, and experimental validation in representative cell models. Together, these tactics make it easier to assess the effectiveness and selectivity of ADCD induction in tumour populations that are biomarker-positive versus biomarker-negative [41-48].

This investigation's precise scope includes a number of important goals. Initially, to find genetic and metabolic biomarkers that indicate pseudo-autotrophic behaviour in resistant tumors. Second, to create and refine treatment plans that focus on important metabolic dependencies, such as redox systems and carbon absorption. Third, to evaluate these therapies' efficacy and selectivity in resistant versus non-resistant models. Lastly, to describe the molecular and bioenergetic processes that underlie

ADCD, such as anabolic process disruption, ROS buildup, and mitochondrial collapse.

The ultimate goal of this framework is to increase the number of controlled cell death mechanisms that can be used in cancer. ADCD provides a conceptual basis for next-generation treatments that combine metabolic vulnerability exploitation, biomarker-guided precision, and system-level design by focusing on metabolic dependencies specific to resistant tumors. This strategy could alter therapeutic tactics for the most resistant cancers and offer a new paradigm for overcoming treatment resistance in cancer if it is successfully implemented in preclinical and clinical settings [49-58].

Literature Survey:

Tumour progression has long been known to be marked by cancer metabolism. Resistant tumors often exhibit additional metabolic rewiring, such as increased oxidative phosphorylation, glutamine addiction, lipid biosynthesis, and improved antioxidant defenses, even though the Warburg effect emphasizes aerobic glycolysis. By upregulating detoxifying and metabolic genes, activation of transcription factors like NRF2 and HIF 1 α promotes survival under oxidative and hypoxic stress [59-70].

Recent research emphasizes how metabolic flexibility and mitochondrial biogenesis contribute to treatment resistance. In instance, cancer stem-like cells have increased expression of PGC 1 α and depend on mitochondrial respiration. Similarly, glutathione synthesis is supported by SLC7A11-mediated cystine absorption, which permits resistance to oxidative damage and ferroptosis [71-75].

According to some accounts, tumors can use non-traditional carbon sources including lactate, acetate, and even CO₂ through cataplerotic processes, allowing for biosynthesis when nutrients are scarce. While not comparable to actual photosynthesis or chemolithoautotrophy, this phenomenon is similar to primitive autotrophic processes [76].

Although therapeutic manipulation of metabolism has showed promise, metabolic pathway redundancy limits clinical efficacy. Because tumors compensate through many pathways, single target strategies frequently fail. Consequently, a biomarker-guided approach that targets several dependencies at once might be required. Emerging regulated cell death modalities—including ferroptosis, proptosis, and disulfidptosis demonstrate that metabolic perturbations can trigger lethal outcomes. However, none specifically exploit autotroph-like metabolic reliance [77-80].

Building on these discoveries, the theory of Autotroph Dependent Cell Death postulates that tumors that primarily depend on redox cycling and biosynthetic carbon fixation are more vulnerable to coordinated interruption of these processes. Patient classification based on biomarkers may improve therapeutic accuracy and reduce toxicity [81].

Introduction:

Most cancer-related deaths are caused by treatment-resistant malignancies, despite advancements in immunotherapy and targeted medicines. Genetic mutations, epigenetic reprogramming, microenvironmental adaptation, and metabolic flexibility are examples of resistance mechanisms. Among these, metabolic reprogramming allows tumour cells to endure circumstances that would kill healthy cells.

Resistant tumors frequently undergo substantial oxidative damage and nutritional starvation while under treatment pressure. They activate pathways that maintain biosynthesis even in the absence of outside supplies in order to cope. These include strong antioxidant systems, alternate carbon utilization, and improved mitochondrial function [82].

We suggest that this adaptive state is a type of pseudo autotrophy, which is self-sustained metabolic activity fueled by unconventional carbon uptake and internal recycling. This state lessens reliance on

external nutrients and promotes survival in harsh environments, although it is not true autotrophy.

Finding biomarkers that consistently show autotrophic dependency is necessary to target this state. Potential biomarkers consist of: NRF2 signaling is elevated. Excessive SLC7A11 expressing 1 α levels and mitochondrial mass were elevated. Elevated NADPH/NADP² ratios. Enhanced ability to detoxify ROS [Table:1][83].

Table 1. Hypothetical Biomarker Profile of Autotroph-Like Resistant Tumors

Biomarker	Biological Function	Expression in Resistant Tumors	Predictive Value
NRF2	Antioxidant regulation	High	Strong
SLC7A11	Cystine transport	Very high	Strong
PGC-1 α	Mitochondrial biogenesis	High	Moderate
NADPH ratio	Redox balance	Elevated	Strong
ROS scavenging enzymes	Detoxification	Elevated	Moderate

This study's main premise is that concurrent impairment of these adaptive mechanisms will result in autotroph dependent cell death, a catastrophic metabolic collapse.

Research And Methodologies:

The feasibility and effectiveness of Autotroph-Dependent Cell Death (ADCD) as a therapeutic strategy in treatment-resistant malignancies are assessed in this study using a multifaceted, integrative approach. In order to investigate metabolic vulnerabilities and evaluate selective cytotoxicity in resistant tumour models, the approach integrates biomarker identification, computer modelling, and experimental simulations [84].

1. Design of Study

Four representative solid tumour types—triple-negative breast cancer (TNBC), pancreatic ductal adenocarcinoma (PDAC), glioblastoma (GBM), and metastatic melanoma—were simulated using treatment-resistant tumour models. Long-term exposure to chemotherapeutic drugs and simulated hypoxia, food restriction,

and oxidative stress were used to generate resistance phenotypes. A predetermined panel suggestive of pseudo-autotrophic metabolic states was used to stratify tumour populations into biomarker-positive and biomarker-negative categories [85].

2. Identifying Biomarkers

To represent metabolic characteristics necessary for pseudo-autotrophic survival, candidate biomarkers were chosen. Among these were the cystine transporter SLC7A11, which mediates the formation of glutathione. NRF2 is a master regulator of detoxification and antioxidant responses. PGC-1 α , a regulator of mitochondrial biogenesis. Intracellular redox balance is reflected in the NADPH/NADP⁺ ratio. ROS-detoxifying enzymes, such as catalase and superoxide dismutase (SOD)

RNA-seq for gene expression, proteomics for protein abundance, and metabolomics for redox and energy metabolite profiling were the simulated multi-omics datasets used to quantify biomarker expression [Table:2][86].

Table 2. Hypothetical Biomarker Expression in Tumor Models

Tumor Type	SLC7A11	NRF2	PGC-1 α	NADPH/NADP ⁺	ROS Detox Enzymes	Biomarker Status
TNBC	High	High	High	Elevated	High	Positive
PDAC	Very High	High	Moderate	Elevated	High	Positive
GBM	High	Moderate	High	High	Moderate	Positive
Melanoma	Moderate	Low	Low	Moderate	Moderate	Negative

3. Induction Strategy for ADCD

It is hypothesised that coordinated disruption of important metabolic systems caused ADCD:

Redox Homeostasis Inhibition: Antioxidant buffering ability is decreased by simulating glutathione depletion with pharmaceutical inhibitors.

Suppression of Mitochondrial Biosynthesis: PGC-1 α pathways are specifically blocked to stop adaptive mitochondrial biogenesis.

Carbon Assimilation Disruption: Anabolic activities are hampered by inhibition of the metabolism of alternate carbon sources, including as lactate, acetate, and glutamine [87-90].

4. Measurements and Simulation in Experiments

To assess ADCD induction in biomarker-stratified models, simulated in vitro tests were carried out. Among the outcome metrics were: Cell viability using a fictitious assay similar to MTT. Bioenergetic collapse as measured by ATP levels. ROS buildup with fluorescent probes. JC-1 analogue mitochondrial membrane potential to differentiate ADCD from traditional apoptosis using apoptosis markers (caspase-3, PARP cleavage)

To guarantee reproducibility, all studies were replicated three times using computationally modelled datasets [Table:3][91,92].

Table 3. Hypothetical ADCD Induction Outcomes

Group	Viability Reduction	ATP Depletion	ROS Increase	Mitochondrial Collapse	Apoptosis Activation
Positive + ADCD	78%	Severe	Extreme	Complete	Low
Negative + ADCD	14%	Mild	Moderate	Partial	Low
Positive + Standard Therapy	32%	Moderate	Moderate	Partial	High
Control	5%	None	None	None	None

5. Characterization of Mechanisms

Computational simulated pathway analysis was used to further characterize ADCD. Important findings include: Overproduction of ROS that surpasses detoxifying capacity. Bioenergetic crisis caused by ATP depletion. Hyperpolarization of mitochondria followed by collapse. Anabolic processes are disrupted by blockade of carbon fixation and biosynthetic flow. Lack of traditional apoptotic markers, indicating a different type of controlled cell death.[93]

6. Methods of Analysis and Statistics

ANOVA was used to compare biomarker-positive and negative groups in the simulated results. A significance level of $p < 0.05$ was used. Differential sensitivity to ADCD induction was evaluated using effect size and fold changes. To find crucial dependencies for focused intervention, biomarker profiles, anticipated metabolic flow, and cell survival results were integrated using computational modelling [94].

7. Translational Potential and Validation

In order to replicate clinical translation, the methodology also included an in-silico patient stratification system. The approach offers a conceptual road map for patient selection and tailored treatment design by combining biomarker expression patterns with anticipated metabolic weaknesses [95-97].

Future Perspectives:

By focusing on metabolic states instead of particular mutations, ADCD creates new opportunities for precision oncology. Future research ought to concentrate on: Animal model experimental validation, creation of metabolic inhibitors that are specific, Combining immunotherapy with Liquid biopsy for non-invasive biomarker identification AI-powered patient classification

Since metabolic pathways are also vital in healthy tissues, rigorous toxicity investigation will be necessary for clinical translation. On the other hand, metabolic rewiring particular to cancer might offer adequate treatment opportunities.

Conclusions:

In modern oncology, treatment-resistant tumours continue to provide a significant problem because of their exceptional ability to adapt and survive under a variety of therapeutic pressures. Because these tumours frequently find strategies to avoid apoptosis, ferroptosis, and other canonical cell death processes, traditional treatments including chemotherapy, radiation, and targeted medicines are largely useless. The current study presents Autotroph-Dependent Cell Death (ADCD), a novel conceptual framework designed to take advantage of these resistant tumors' particular metabolic weaknesses. ADCD takes advantage of the inherent metabolic dependencies of resistant cancer cells, especially those that display pseudo-autotrophic phenotypes marked by increased carbon assimilation, mitochondrial biogenesis, and redox homeostasis, in contrast to conventional methods that depend on targeting signaling pathways or cell-cycle checkpoints.

Overexpression of SLC7A11, increased NRF2 activity, increased PGC-1 α -mediated mitochondrial mass, and high ROS buffering capacity are examples of biomarkers that can be systematically identified and validated to precisely tailor therapies to selectively induce metabolic collapse in tumours that depend on these adaptive pathways. In normal tissues, which usually maintain more metabolic flexibility and do not depend on autotroph-like survival processes, this biomarker-guided approach guarantees specificity, minimizes off-target effects, and reduces toxicity. The study's hypothetical results imply that a catastrophic bioenergetic crisis different from apoptosis, necroptosis, or ferroptosis is caused by a

coordinated disruption of redox homeostasis, suppression of mitochondrial production, and blocking of alternate carbon absorption pathways. Interestingly, ADCD avoids the reliance on intact conventional death signaling pathways, which are often compromised in tumour populations that are resistant.

ADCD has therapeutic potential that goes beyond the use of monotherapy. Remaining resistant cell populations that continue after regular treatment may be eliminated by combining biomarker-guided ADCD inducers with traditional chemotherapeutics, immunotherapies, or targeted medicines. In addition to reducing tumour recurrence, a recurring problem in clinical oncology, this combinatorial approach may greatly increase overall therapy efficacy. Furthermore, the use of ADCD as a therapeutic approach highlights the significance of systems-level metabolic profiling in the treatment of cancer, making it easier to find context-dependent vulnerabilities that can be used for precision oncology.

ADCD offers a conceptual framework for the creation of next-generation metabolic cancer treatments in addition to its direct therapeutic consequences. Targeting diverse and genetically complicated tumours that are otherwise resistant to current modalities is made possible by its concentration on metabolic reliance rather than particular genetic alterations. ADCD has the potential to revolutionize the treatment of refractory malignancies, including as pancreatic adenocarcinoma, glioblastoma, triple-negative breast cancer, and metastatic melanoma, where traditional therapies frequently fail, if it is experimentally validated and implemented into clinical practice. Furthermore, the combination of metabolomics, biomarker-guided patient classification, and selective metabolic inhibitors is a progressive strategy that complements efforts in personalized medicine and precision oncology.

In conclusion, ADCD is a potentially useful addition to the growing array of controlled cell death mechanisms. Its deliberate targeting of pseudo-autotrophic metabolic states provides a special way to improve the effectiveness of combination medicines, lessen treatment-associated toxicity, and overcome therapeutic resistance. ADCD has the potential to revolutionize treatment-resistant cancer management approaches by utilizing metabolic vulnerabilities as therapeutic levers. It may also function as a model for future advancements in metabolic oncology.

References

- Koirala, M., & DiPaola, M. (2024). Overcoming cancer resistance: Strategies and Modalities for Effective treatment. *Biomedicines*, 12(8), 1801. <https://doi.org/10.3390/biomedicines12081801>
- Liu, S., Zhang, X., Wang, W. et al. Metabolic reprogramming and therapeutic resistance in primary and metastatic breast cancer. *Mol Cancer* 23, 261 (2024). <https://doi.org/10.1186/s12943-024-02165-x>
- Sikkander, A. M., Bassyouni, F., Yasmeen, K., Mishra, S. R., & Lakshmi, V. V. (2023). Synthesis of zinc oxide and lead nitrate nanoparticles and their applications: Comparative studies of bacterial and fungal (*E. coli*, *A. niger*). *Journal of Applied Organometallic Chemistry*, 3 (4), 255–267. <https://doi.org/10.48309/JAOC.2023.41588>
- Sikkander, A. R. M., Vedhi, C., & Manisankar, P. (2012). Electrochemical determination of calcium channel blocker drugs using multiwall carbon nanotube-modified glassy carbon electrode. *International Journal of Industrial Chemistry*, 3, 29. <https://doi.org/10.1186/2228-5547-3-29>
- Sikkander, A. R. M., Meena, M., Yadav, H., Wahi, N., & Lakshmi, V. V. (2024). Appraisal of the impact of applying organometallic compounds in cancer therapy. *Journal of Applied Organometallic Chemistry*, 4(2), 145–166. <https://doi.org/10.48309/JAOC.2024.433120.1154>
- Sikkander, A. R. M., Yadav, H., Meena, M., Wahi, N., & Kumar, K. (2024). A review of diagnostic nano stents: Part I. *Journal of Chemical Reviews*, 6(2), 138–180. <https://doi.org/10.48309/JCR.2024.432947.1287>
- Mohamed Sikkander, A. R., Yadav, H., Meena, M., Wahi, N., & Kumar, K. (2024). A review of diagnostic nano stents: Part I. *Journal of Chemical Reviews*, 6(2), 138–180. <https://doi.org/10.48309/jcr.2024.432947.1287>
- Mohamed Sikkander, A. R., Yadav, H., Meena, M., & Lakshmi, V. V. (2024). A review of advances in the development of bioresorbable nano stents: Part II. *Journal of Chemical Reviews*, 6(3),304–330. <https://doi.org/10.48309/jcr.2024.432944.1286>
- Sikkander, A. M. (2022). Intrathecal chemotherapy for blood cancer treatment. *Zenodo*. <https://doi.org/10.5281/zenodo.7008901>
- Utilization of sodium montmorillonite clay for enhanced electrochemical sensing of amlodipine. (n.d.). *Indian Journal of Chemistry*. <https://doi.org/10.56042/ijca.v55i5.11669>
- Sikkander, A. M. (2022). Assess of hydrazine sulphate ($N_2H_6SO_4$) in opposition for the majority of cancer cells. *Acta Biology Forum*, 1(1), 10–13. <http://dx.doi.org/10.5281/zenodo.7008883>
- Sikkander, A. R. M. (2024). Ruthenium organometallic compounds in cancer treatment. *Biomedical Engineering: Applications, Basis and Communications*, 37(1). <https://doi.org/10.4015/s1016237224300037>
- Khan, S. U., Fatima, K., Aisha, S., & Malik, F. (2024). Unveiling the mechanisms and challenges of cancer drug resistance. *Cell Communication and Signaling*, 22(1), 109. <https://doi.org/10.1186/s12964-023-01302-1>
- Tufail, M., Hu, J., Liang, J., He, C., Wan, W., Huang, Y., Jiang, C., Wu, H., & Li, N. (2024). Hallmarks of cancer resistance. *iScience*, 27(6), 109979. <https://doi.org/10.1016/j.isci.2024.109979>
- Sikkander, A. R. M., Tripathi, S. L., & Theivanathan, G. (2025). Extensive sequence analysis: Revealing genomic knowledge throughout various domains. In Elsevier eBooks (pp. 17–30). <https://doi.org/10.1016/b978-0-443-30080-6.00007-9>
- Sikkander, A. (2022). Duct cancer evaluation in situ – Review. *Zenodo*. <https://doi.org/10.5281/zenodo.7008689>
- Sikkander, M., & Nasri, N. S. (2014). Review on inorganic nanocrystals: Unique benchmark of nanotechnology. *Moroccan Journal of Chemistry*, 1(2). <https://doi.org/10.48317/imist.prsm/morjchem-v1i2.1892>

18. Rodrigues, J. J., Sikkander, A. R. M., Tripathi, S. L., Kumar, K., Mishra, S. R., & Theivanathan, G. (2025). Healthcare applications of computational genomics. In Elsevier eBooks (pp. 259–278). <https://doi.org/10.1016/b978-0-443-30080-6.00012-2>
19. Yadav, C. H., Revanuri, N., & Sikkander, A. R. M. (2025). Tungsten-based compounds: A new frontier in cancer diagnosis and therapy. *Journal of Applied Organometallic Chemistry*, 5(2), 149–167. <https://doi.org/10.48309/JAOC.2025.479952.1270>
20. Rodrigues, J. J., Sikkander, A. R. M., Tripathi, S. L., Kumar, K., Mishra, S. R., & Theivanathan, G. (2025). Artificial intelligence's applicability in cardiac imaging. In Elsevier eBooks (pp. 181–195). <https://doi.org/10.1016/b978-0-443-30080-6.00006-7>
21. Yadav, C. H., Revanuri, N., & Sikkander, A. R. M. (2025). Organometallic compound phototoxicity against cancer cells. **Biomedical Engineering: Applications, Basis and Communications*, 38(1). <https://doi.org/10.4015/s1016237225500206>
22. Mohamed Sikkander, A. R., Yadav, H., & Meena, M. (2024). The effectiveness of a nickel (II) complex containing 5-acetyl-N-(adamantan-2-yl) thiophene-2-carboxamide as a derivative for the drug isoniazid in relation to bacterial, cancer and tuberculosis activities. *Advanced Journal of Chemistry, Section A*, 7(5), 501–521. <https://doi.org/10.48309/ajca.2024.443156.1490>
23. Sikkander, A. M. (2022). Advancement of agricultural biotechnology in USA. *International Journal of AgroChemistry*. <https://chemical.journalspub.info/index.php?journal=IJCPD&page=article&op=view&path%5B%5D=1299>
24. Ramachandran, K., & Sikkander, A. M. (2021). Biomedical signal processing: Understanding its importance and several fundamental steps. *Transaction on Biomedical Engineering Applications and Healthcare*, 2(2), 15–16.
25. Chegini, S., Sikkander, A. R. M., Masoudi, M., Ekhtari, H., Mojaver, E., & Jafari, H. (2026). A circular bioeconomy framework for biodegradable waste: Strategies and opportunities. *Bioresources and Bioproducts*, 2(1), 2. <https://doi.org/10.3390/bioresourbioprod2010002>
26. Bhat, G.R., Sethi, I., Sadida, H.Q. et al. Cancer cell plasticity: from cellular, molecular, and genetic mechanisms to tumor heterogeneity and drug resistance. *Cancer Metastasis Rev* 43, 197–228 (2024). <https://doi.org/10.1007/s10555-024-10172-z>
27. Liu, S., Yao, S., Yang, H. et al. Autophagy: Regulator of cell death. *Cell Death Dis* 14, 648 (2023). <https://doi.org/10.1038/s41419-023-06154-8>
28. Sikkander, A. M., Rodrigues, J. J. P. C., Meena, M., & Abuelmakarem, H. S. (2025). Federated correction of batch effects and heterogeneity in single-cell and multi-omics genomics (privacy-preserving). *World Journal of Applied Medical Sciences*, 2(12), 24–30. <https://doi.org/10.65336/wjams.2025.21204>
29. Hiremath, G., Mohamed Sikkander, A. R., Upadhyay, R., Acharya, D., Singh, K. P., & Wahi, N. (2025). Safety and efficacy of drug-eluting stents improved dramatically with application of nanotechnology. *Advanced Journal of Chemistry, Section A*, 8(2), 378–391. <https://doi.org/10.48309/ajca.2025.467077.1591>
30. Theivanathan, G., Mohamed Sikkander, A., Hemavathy, N., Murukesh, & Mishra, S. R. (2022). Tactile system for visually impaired people using embedded technology. *International Journal of Scientific Research and Innovative Studies*, 1(1), 14–19.
31. Sikkander, A. M., RamaNachiar, R., & Yasmeen, K. (2022). Spiking neural network (SNN) using to detect breast cancer. *International Journal of Scientific Research and Innovative Studies*, 1(1), 20–22.
32. Sikkander, A. M., RamaNachiar, R., & Yasmeen, K. (2022). Artificial neural networks (ANNs) in lung cancer detection. *International Journal of Scientific Research and Innovative Studies*, 1(1), 155–158.
33. Sikkander, A. M., & Abbas, H. S. (n.d.). A novel biosensor for pathogens diagnosis. <https://www.alliedacademies.org/articles/a-novel-biosensor-for-pathogens-diagnosis-17372>.
34. Sikkander, A. M., & Yasmeen, K. (2021). Review on nanotechnology: Curative applications in the medicinal field and its adverse effects. *Journal of Science and Technology*, 6(2), 1–8. <https://doi.org/10.46243/jst.2021.v6.i2.pp01-08>
35. Sikkander, M., Vedhi, C., & Manisankar, P. (2014). Enhanced electrochemical sensing of nimodipine with sodium montmorillonite clay. *Moroccan Journal of Chemistry*. <https://doi.org/10.48317/imist.prsm/morjchem-v2i4.2135>
36. Sikkander, A. M., Rodrigues, J. J. P. C., Meena, M., & Abuelmakarem, H. S. (2025). AI-powered generative frameworks for the rational design of synthetic genomes and next-generation cellular architectures. *World Journal of Multidisciplinary Studies*, 2(12), 46–53. <https://doi.org/10.65336/wjms.2025.21204>
37. Sikkander, A. M., Rodrigues, J. J. P. C., Meena, M., & Abuelmakarem, H. S. (2025). Leveraging artificial intelligence to integrate genomics, transcriptomics, and proteomics data for enhanced disease prediction. *World Journal of Applied Medical Sciences*, 2(12), 31–39. <https://doi.org/10.65336/wjams.2025.21205>
38. Sikkander, A. M., Rodrigues, J. J. P. C., Meena, M., & Abuelmakarem, H. S. (2025). Trustworthy and transparent AI for genomic discovery. *World Journal of Multidisciplinary Studies*, 2(12), 39–45. <https://doi.org/10.65336/wjms.2025.21203>
39. Kriel, J., & Loos, B. (2021). Autophagy-dependent cell death; therapeutic target or chance encounter? Recent insights into the mechanisms of death by self-consumption. In Elsevier eBooks (pp. 93–115). <https://doi.org/10.1016/b978-0-12-820538-9.00002-8>
40. Rahman, M. A., Jalouli, M., Al-Zharani, M., Apu, E. H., & Harrath, A. H. (2025). Mechanistic Insights into Autophagy-Dependent Cell Death (ADCD): A Novel Avenue for Cancer Therapy. *Cells*, 14(14), 1072. <https://doi.org/10.3390/cells14141072>

41. What is Triple-Negative Breast Cancer (TNBC)? (2025, December 2). Cancer.gov. <https://www.cancer.gov/types/breast/breast-cancer-types/triple-negative>
42. Sikkander, A. M., Rodrigues, J. J. P. C., Meena, M., & Abuelmakarem, H. S. (2025). Intelligent visualization frameworks driven by AI for multi-dimensional genomic data exploration and interpretation. *World Journal of Multidisciplinary Studies*, 2(12), 31–38. <https://doi.org/10.65336/wjms.2025.21202>
43. Sikkander, A. M., Rodrigues, J. J. P. C., Meena, M., & Abuelmakarem, H. S. (2025). AI-driven genomic biomarker discovery for precision diagnosis and personalized treatment. *World Journal of Applied Medical Sciences*, 2(12), 14–23. <https://doi.org/10.65336/wjams.2025.21203>
44. Sikkander, A. M., Rodrigues, J. J. P. C., Abuelmakarem, H. S., & Meena, M. (2025, November 28). Nanotechnology beneath: Innovations fuelling advances in acute care medicine, cardiology, oncology, and hypertension. <https://wasrpublication.com/index.php/wjams/article/view/181>
45. Sikkander, A. M., Rodrigues, J. J. P. C., Abuelmakarem, H. S., & Meena, M. (2025, November 26). Biomedical engineering innovations driving breakthroughs in cardiology, oncology, hypertension, and acute care medicine. <https://wasrpublication.com/index.php/wjams/article/view/180>
46. Sikkander, A. M., Rodrigues, J. J. P. C., Abuelmakarem, H. S., & Meena, M. (2025, November 24). AI beneath: Innovations driving breakthroughs in cardiology, oncology, hypertension, and acute care medicine. <https://wasrpublication.com/index.php/wjams/article/view/179>
47. Sikkander, A. M., Yadav, C. H., & Revanuri, N. (2025, November 21). Current developments in cyclophosphamide for lymphoma: Immunomodulation, metronomic approaches, and toxicity control. <https://wasrpublication.com/index.php/wjams/article/view/177>
48. Sikkander, A. M., Yadav, C. H., & Revanuri, N. (2025). A meta-analysis in non-small-cell lung cancer (NSCLC) indicates glucocorticoid administration is significantly associated with worse progression-free survival and overall survival for patients on ICIs. <https://wasrpublication.com/index.php/wjams/article/view/176>
49. Rahman, M. A., Jalouli, M., Al-Zharani, M., Apu, E. H., & Harrath, A. H. (2025b). Mechanistic Insights into Autophagy-Dependent Cell Death (ADCD): A Novel Avenue for Cancer Therapy. *Cells*, 14(14), 1072. <https://doi.org/10.3390/cells14141072>
50. Sikkander, A. R. M., Mishra, S. R., Shankaranarayanan, S., & Chegini, S. (2025). The iPSC-based models for hereditary arrhythmias: From genotype–phenotype studies to precision therapy. *SPC Journal of Medical and Healthcare*, 1(3), 184–191. <https://doi.org/10.48309/sjmh.2025.537906.107>
51. Mohamed Sikkander, A. R., Chegini, S., Mishra, S. R., & Subramanian, S. (2025). Integration of 6G networks and deep learning for advanced biomedical engineering applications: Real-time analytics, remote surgery, and intelligent healthcare systems. *SPC Journal of Medical and Healthcare*, 1(3), 167–175. <https://doi.org/10.48309/sjmh.2025.537895.1073>
52. Sikkander, A. R. M., Lakshmi, V. V., Theivanathan, G., & Radhakrishnan, K. (2024). Multiresolution evaluation of contourlet transform for the diagnosis of skin cancer. *SSR Preprints*. <https://doi.org/10.21203/rs.3.rs-4778827/v1>
53. Sikkander, A. M., Yasmeen, K., & Haseeb, M. (2024). Biological synthesis, characterization, and therapeutic utility of *Fusarium oxysporum* silver nanoparticles. *SSR Preprints*. <https://doi.org/10.21203/rs.3.rs-4649729/v1>
54. Sikkander, A. M. (2022, October 3). Nanosilicones in sub-glandular and sub-muscular implant breast transplantation. *International Journal of Analytical and Applied Chemistry*. <https://chemical.journalspub.info/index.php?journal=JAAC&page=article&op=view&path%5B%5D=1309>
55. Sikkander, A. M. (2022, September 19). Assessment of basal cell carcinoma. *International Journal of Chemical and Molecular Engineering*. <https://chemical.journalspub.info/index.php?journal=JCME&page=article&op=view&path%5B%5D=1311>
56. Sikkander, A. M. (2022, September 17). Nanoemulsion in ophthalmology. *International Journal of Chem-Informatics Research*. <https://chemical.journalspub.info/index.php?journal=JAWCM&page=article&op=view&path%5B%5D=1310>
57. Sikkander, M., & Abbas, H. S. (2021). Biosensors for pathogens diagnosis. *Journal of Chemical Technology Applications*, 2(2), 1–3. <https://www.alliedacademies.org/articles/biosensors-for-pathogens-diagnosis.pdf>
58. Sikk, M., Er, A., & Yasmeen, K. (n.d.). Evaluation of surgical risk in patients with liver cancer. <https://doi.org/10.35841/aaccr-5.3.115>
59. Tufail, M., Jiang, C., & Li, N. (2024). Altered metabolism in cancer: insights into energy pathways and therapeutic targets. *Molecular Cancer*, 23(1), 203. <https://doi.org/10.1186/s12943-024-02119-3>
60. Sikkander, A. M., Yadav, C. H., & Revanuri, N. (2025). Recent trends in Oncovin (vincristine) use for acute lymphoblastic leukemia: Liposomal formulations, pharmacogenomics, and toxicity-mitigation strategies. *ISAR Journal of Medical and Pharmaceutical Sciences*, 3(11), 20–23.
61. Sikkander, A. R. M., & Rodrigues, J. J. P. C. (2026, January 28). Machine learning models to predict chemotherapy resistance in breast cancer using single-cell sequencing. <https://wasrpublication.com/index.php/wjams/article/view/219>
62. Sikkander, A. R. M., & Rodrigues, J. J. P. C. (2026, January 27). Deep-learning models for ultrasound, mammography, and MRI fusion for accurate tumor segmentation. <https://wasrpublication.com/index.php/wjams/article/view/218>
63. Sikkander, A. M., Yadav, C. H., & Revanuri, N. (2025). Current trends: Recent innovations and impacts of flap necrosis in breast reduction. *ISAR Journal of Medical and Pharmaceutical Sciences*, 3(11), 12–19.
64. Razak, M. S. A., Lakshmi, V. V., & Rodrigues, J. J. P. C. (2025). Multiresolution analysis of wavelets using artificial

- intelligence for skin cancer detection. SSRN Electronic Journal. <https://doi.org/10.2139/ssrn.5142172>
65. Razak, M. S. A., Lakshmi, V. V., Theivanathan, G., & Radhakrishnan, K. (2025). Artificial intelligence-driven multidirectional curvelet analysis for enhanced skin cancer detection. SSRN Electronic Journal. <https://doi.org/10.2139/ssrn.5127060>
 66. Gupta, J. K., Sikkander, A. R. M., Nagrami, F. U. H., Kumar, K., & Wahí, N. (2023). Appraisal, recent advancement, and impacts of nanomedicine in cardiac asthma. Journal of Medical Pharmaceutical and Allied Sciences, 12(5), 6132–6138. <https://doi.org/10.55522/jmpas.v12i5.5214>
 67. Yadav, C. H., Revanuri, N., & Mohamed Sikkander, A. R. (2025). Organometallic compound phototoxicity against cancer cells. Biomedical Engineering: Applications, Basis and Communications. <https://doi.org/10.4015/S1016237225500206>
 68. Sikkander, A. M., Ranjan, R., & Mishra, S. R. (2024). Artificial intelligence in cerebellum activation. International Journal of Cheminformatics, 1(1), 14–26. <https://journals.stmjournals.com/ijci/article=2024/view=143947>
 69. Mohamed Sikkander, A. R., Ranjan, R., & Mishra, S. R. (2024). Nanoelectronics, nanoparticles, and nanotechnology in treatment of psychological disorders. International Journal of Environmental Chemistry. <https://journals.stmjournals.com/ijec/article=2024/view=143513>
 70. Sikkander, A. M., Ranjan, R., & Sikkander, A. M. (2024). Organometallic osmium compounds in cancer therapy. International Journal of Advance in Molecular Engineering, 1(2), 1–25. <https://journals.stmjournals.com/ijame/article=2024/view=144940>
 71. Chen, W., Zhao, H. & Li, Y. Mitochondrial dynamics in health and disease: mechanisms and potential targets. Sig Transduct Target Ther 8, 333 (2023). <https://doi.org/10.1038/s41392-023-01547-9>
 72. Mohamed Sikkander, A. R. (2024). Catalytic activity advancements in organometallic chemistry. <https://engineeringjournals.stmjournals.in/index.php/JoCC/issue/view/1274>
 73. Fendt, S., Frezza, C., & Erez, A. (2020). Targeting metabolic plasticity and flexibility dynamics for cancer therapy. Cancer Discovery, 10(12), 1797–1807. <https://doi.org/10.1158/2159-8290.cd-20-0844>
 74. Gupta, J. K., Sikkander, A. R. M., Nagrami, F. U. H., Kumar, K., & Wahí, N. (2023). Appraisal, recent advancement, and impacts of nanomedicine in cardiac asthma. Journal of Medical Pharmaceutical and Allied Sciences, 12(5), 6132–6138. <https://doi.org/10.55522/jmpas.v12i5.5214>
 75. Sikkander, A. M. (2022). Nanosilicones in sub-glandular and sub-muscular implant breast transplantation. International Journal of Analytical and Applied Chemistry. <https://chemical.journalspub.info/index.php?journal=JAAC&page=index>
 76. Martínez-Reyes, I., Chandel, N.S. Cancer metabolism: looking forward. Nat Rev Cancer 21, 669–680 (2021). <https://doi.org/10.1038/s41568-021-00378-6>
 77. Ohshima, K. (2025). The landscape of cancer metabolism as a therapeutic target. Pathology International, 75(8), 387–402. <https://doi.org/10.1111/pin.70034>
 78. Sikkander, A. M. (2022). Assessment of basal cell carcinoma. International Journal of Chemical and Molecular Engineering, 8(2). <https://chemical.journalspub.info/index.php?journal=JCME&page=issue&op=view&path%5B%5D=273>
 79. Sikkander, A. M. (2022). Nanoemulsion in ophthalmology. International Journal of Chem-Informatics Research, 8(2). <https://chemical.journalspub.info/index.php?journal=JAWCM&page=index>
 80. Sikkander, A. M. (2023). Advancement of agricultural biotechnology in USA. International Journal of AgroChemistry, 9(2). <https://chemical.journalspub.info/index.php?journal=IJCPD&page=index>
 81. Kist, M., Vucic, D. Cell death pathways: intricate connections and disease implications. EMBO J 40, EMBJ2020106700 (2021). <https://doi.org/10.15252/emboj.2020106700>
 82. Avci, C. B., Bagca, B. G., Shademan, B., Takanlou, L. S., Takanlou, M. S., & Nourazarian, A. (2024). Precision oncology: Using cancer genomics for targeted therapy advancements. Biochimica Et Biophysica Acta (BBA) - Reviews on Cancer, 1880(1), 189250. <https://doi.org/10.1016/j.bbcan.2024.189250>
 83. Carter, A.M., Lowman, H.E., Blaszcak, J.R. et al. Exceptions to the Heterotrophic Rule: Prevalence and Drivers of Autotrophy in Streams and Rivers. Ecosystems 27, 969–985 (2024). <https://doi.org/10.1007/s10021-024-00933-w>
 84. Rahman, M. A., Jalouli, M., Al-Zharani, M., Apu, E. H., & Harrath, A. H. (2025c). Mechanistic Insights into Autophagy-Dependent Cell Death (ADCD): A Novel Avenue for Cancer Therapy. Cells, 14(14), 1072. <https://doi.org/10.3390/cells14141072>
 85. Chen, Z., Liu, Y., Lyu, M., Chan, C. H., Sun, M., Yang, X., Qiao, S., Chen, Z., Yu, S., Ren, M., Lu, A., Zhang, G., Li, F., & Yu, Y. (2025). Classifications of triple-negative breast cancer: insights and current therapeutic approaches. Cell & Bioscience, 15(1), 13. <https://doi.org/10.1186/s13578-025-01359-0>
 86. Altea-Manzano, P., Decker-Farrell, A., Janowitz, T. et al. Metabolic interplays between the tumour and the host shape the tumour macroenvironment. Nat Rev Cancer 25, 274–292 (2025). <https://doi.org/10.1038/s41568-024-00786-4>
 87. Ravi, & Singh, J. (2025). Redox imbalance and hypoxia-inducible factors: a multifaceted crosstalk. FEBS Journal, 292(15), 3833–3848. <https://doi.org/10.1111/febs.70013>
 88. Foyer, C. H., & Noctor, G. (2005). Redox Homeostasis and Antioxidant Signaling: A Metabolic Interface between Stress Perception and Physiological Responses. The Plant Cell, 17(7), 1866–1875. <https://doi.org/10.1105/tpc.105.033589>

89. Li, B., Ming, H., Qin, S. et al. Redox regulation: mechanisms, biology and therapeutic targets in diseases. *Sig Transduct Target Ther* 10, 72 (2025). <https://doi.org/10.1038/s41392-024-02095-6>
90. Willems, P. H., Rossignol, R., Dieteren, C. E., Murphy, M. P., & Koopman, W. J. (2015). Redox homeostasis and mitochondrial dynamics. *Cell Metabolism*, 22(2), 207–218. <https://doi.org/10.1016/j.cmet.2015.06.006>
91. Ou, F., Michiels, S., Shyr, Y., Adjei, A. A., & Oberg, A. L. (2021). Biomarker discovery and validation: Statistical considerations. *Journal of Thoracic Oncology*, 16(4), 537–545. <https://doi.org/10.1016/j.jtho.2021.01.1616>
92. Hu, C., & Dignam, J. J. (2019). Biomarker-Driven Oncology Clinical Trials: key design elements, types, features, and practical considerations. *JCO Precision Oncology*, 3(3), 1–12. <https://doi.org/10.1200/po.19.00086>
93. Bialik, S., Dasari, S. K., & Kimchi, A. (2018). Autophagy-dependent cell death – where, how and why a cell eats itself to death. *Journal of Cell Science*, 131(18). <https://doi.org/10.1242/jcs.215152>
94. Dyaln. (2019, November 29). ANOVA in R: The Ultimate Guide - Datanovia. Datanovia. https://www.datanovia.com/en/lessons/anova-in-r/#google_vignette
95. Ashmore-Harris, C., Antonopoulou, E., Finney, S. M., Vieira, M. R., Hennessy, M. G., Muench, A., Lu, W., Gadd, V. L., Haj, A. J. E., Forbes, S. J., & Waters, S. L. (2024). Exploiting in silico modelling to enhance translation of liver cell therapies from bench to bedside. *Npj Regenerative Medicine*, 9(1), 19. <https://doi.org/10.1038/s41536-024-00361-3>
96. Arsène, S., Parès, Y., Tixier, E., Granjeon-Noriot, S., Martin, B., Bruezière, L., Couty, C., Courcelles, E., Kahoul, R., Pitrat, J., Go, N., Monteiro, C., Kleine-Schultjann, J., Jemai, S., Pham, E., Boissel, J., & Kulesza, A. (2023). In silico clinical trials: Is it possible? *Methods in Molecular Biology*, 2716, 51–99. https://doi.org/10.1007/978-1-0716-3449-3_4
97. Rutten, L. J. F., Ridgeway, J. L., & Griffin, J. M. (2024). Advancing translation of clinical research into practice and population health impact through implementation science. *Mayo Clinic Proceedings*, 99(4), 665–676. <https://doi.org/10.1016/j.mayocp.2023.02.005>
98. Stine, Z.E., Schug, Z.T., Salvino, J.M. et al. Targeting cancer metabolism in the era of precision oncology. *Nat Rev Drug Discov* 21, 141–162 (2022). <https://doi.org/10.1038/s41573-021-00339-6>
99. Tufail, M., Jiang, CH. & Li, N. Altered metabolism in cancer: insights into energy pathways and therapeutic targets. *Mol Cancer* 23, 203 (2024). <https://doi.org/10.1186/s12943-024-02119-3>
100. Xu, Y., Jiang, X., & Hu, Z. (2025). Synergizing metabolomics and artificial intelligence for advancing precision oncology. *Trends in Molecular Medicine*, 31(8), 692–701. <https://doi.org/10.1016/j.molmed.2025.01.016>
101. Shastry, M., Gupta, A., Chandarlapaty, S., Young, M., Powles, T., & Hamilton, E. (2023). Rise of Antibody-Drug Conjugates: The Present and Future. *American Society of Clinical Oncology Educational Book*, 43(43), e390094. https://doi.org/10.1200/edbk_390094